A POPULATION PHARMACOKINETIC MODEL WITH TIME-DEPENDENT COVARIATES MEASURED WITH ERRORS

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Abstract

We propose a population pharmacokinetic (PK) model with time-dependent covariates measured with errors. This model is used to model S-oxybutynin’s kinetics following an oral administration of Ditropan, and allows the distribution rate to depend on time-dependent covariates blood pressure and heart rate, which are measured with errors. We propose two two-step estimation methods: the second order two-step method with numerical solutions of differential equations (2orderND), and the second order two-step method with closed form approximate solutions of differential equations (2orderAD). The proposed methods are computationally easy and require fitting a linear mixed model at the first step and a nonlinear mixed model at the second step. We apply the proposed methods to the analysis of the Ditropan data, and evaluate their performance using a simulation study. Our results show that the 2orderND method performs well, while the 2orderAD method can yield PK parameter estimators that are subject to considerable biases.
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SUMMARY

We propose a population pharmacokinetic (PK) model with time-dependent covariates measured with errors. This model is used to model S-oxybutynin’s kinetics following an oral administration of Ditropan, and allows the distribution rate to depend on time-dependent covariates blood pressure and heart rate, which are measured with errors. We propose two two-step estimation methods: the second order two-step method with numerical solutions of differential equations (2orderND), and the second order two-step method with closed form approximate solutions of differential equations (2orderAD). The proposed methods are computationally easy and require fitting a linear mixed model at the first step and a nonlinear mixed model at the second step. We apply the proposed methods to the analysis of the Ditropan data, and evaluate their performance using a simulation study. Our results show that the 2orderND method performs well, while the 2orderAD method can yield PK parameter estimators that are subject to considerable biases.

KEY WORDS: Differential equations; Laplace approximation; Measurement error; Nonlinear mixed models; Pharmacokinetics; Two-compartment model.
1 Introduction

1.1 The Ditropan Study

S-oxybutynin, a major active component of Ditropan (ALZA Corp.), is commonly prescribed as an antispasmodic for patients with uninhibited neurogenic or reflex neurogenic bladder. It is used to relieve symptoms associated with voiding, such as urge incontinence, urgency, and frequency of urination arising from overactivity of the detrusor muscle (AHFS Drug Information, 2000).

In a Ditropan drug study sponsored by ALZA Corp., the controlled released drug formulation (once a day) was compared to the regular formulation (three doses a day) to establish the bioequivalence. In this paper, we focus on the pharmacokinetics of S-oxybutynin based on the three dose data. This study involved 40 healthy subjects, and the three doses were given 8 hours apart. Each subject had 25 plasma measures over 48 hours. Figure 1(d) shows individual S-oxybutynin plasma concentration time profiles. In addition to the plasma concentration of S-oxybutynin, diastolic blood pressure (DBP), systolic blood pressure (SBP) and heart rate (HR) were measured 9 times repeatedly during the 48 hour window for each subject, and the times these covariates were measured were different from those of the plasma measures. The time profiles of DBP, SBP and HR are given in Figures 1(a)-(c). An objective of this study was to develop a pharmacokinetic (PK) model for the S-oxybutynin plasma concentration by modeling the PK parameters as functions of the time-varying covariates blood pressure and heart rate, which were measured with error. We present in Section 1.2 some preliminary analysis results and provide biological motivations for such a model.

1.2 Preliminary Analysis of the Ditropan Data and Motivation for the Proposed Research

Following an oral administration of S-oxybutynin, absorption from the gut is rapid, and it is metabolized extensively by cytochrome P450 3A4 in the liver and the gut wall. Douchamps
et al. (1988) conducted a one oral dose study and found that S-oxybutynin followed a two-compartment model with linear pharmacokinetics. Li et al. (2002) analyzed the three dose Ditropan data, and found that both the dose-dependent maximum concentration ($C_{max}$) and the dose-dependent area under the concentration ($AUC$) of S-oxybutynin were non-decreasing functions of the three consecutive doses, suggesting that some pharmacokinetic parameters were dose dependent. Therefore, we started our pharmacokinetics analysis from a three separate two-compartment model corresponding to the three doses. Specifically, we assumed that the PK parameters were constant during each dosing period and different among the three dosing periods.

Denote by $t_i$ the time the dose $l$ is given ($l = 1, 2, 3$). For the Ditropan study, $(t_1, t_2, t_3) = (0, 8, 16)$. Let $\{A_{1,i}(t - t_i), A_{2,i}(t - t_i), A_{3,i}(t - t_i)\}$ be the amount of drug in gut, blood and tissue at time $t$ for subject $i$ ($i = 1, \ldots, 40$) from dose $l$ ($l = 1, 2, 3$) respectively, where $A_{k,i}(t - t_i) = A_{k,i}(t - t_i) (k = 1, 2, 3)$ if $t > t_i$ and $0$ if $t \leq t_i$ except $A_{1,i}(0) = 1$. The two-compartment model for dose $l$ can be written as the following differential equations

\[
\frac{dA_{1,i}(t - t_i)}{dt} = -k_{a,i}A_{1,i}(t - t_i)
\]

\[
\frac{dA_{2,i}(t - t_i)}{dt} = k_{a,i}A_{1,i}(t - t_i) - (k_{e,i} + k_{23,i})A_{2,i}(t - t_i) + k_{32,i}A_{3,i}(t - t_i)
\]

\[
\frac{dA_{3,i}(t - t_i)}{dt} = k_{23,i}A_{2,i}(t - t_i) - k_{32,i}A_{3,i}(t - t_i),
\]

where $k_{a,i}$ is the absorption rate, $k_{e,i}$ is the elimination rate, and $k_{23,i}$ and $k_{32,i}$ are the distribution rates for subject $i$ given dose $l$. Note that we here assume that the PK parameters are constant within each dose period and different between the three doses. One can show that under this assumption, the above system of differential equations (1) has a closed form solution. Specifically, we have

\[
A_{2,i}(t - t_i) = \frac{k_{a,i}(k_{23,i} - k_{a,i})e^{-k_{a,i}x(t-t_i)}}{\lambda_{1,i}(\lambda_{2,i} - k_{a,i})} + \frac{k_{a,i}(k_{32,i} - \lambda_{1,i})e^{-\lambda_{1,i}x(t-t_i)}}{\lambda_{2,i}(\lambda_{1,i} - \lambda_{1,i})}
\]

\[
+ \frac{k_{a,i}(k_{23,i} - \lambda_{2,i})e^{-\lambda_{2,i}x(t-t_i)}}{\lambda_{1,i}(\lambda_{2,i} - \lambda_{2,i})} + \frac{k_{a,i}(k_{32,i} - \lambda_{2,i})e^{-\lambda_{2,i}x(t-t_i)}}{\lambda_{1,i}(\lambda_{2,i} - \lambda_{2,i})},
\]
where

\[
\begin{align*}
\lambda_{1,il} &= [k_{e,il} + k_{23,il} + k_{32,il} + \{(k_{e,il} + k_{23,il} + k_{32,il})^2 - 4k_{e,il}k_{32,il}\}]^{1/2}/2 \\
\lambda_{2,il} &= [(k_{e,il} + k_{23,il} + k_{32,il} - \{(k_{e,il} + k_{23,il} + k_{32,il})^2 - 4k_{e,il}k_{32,il}\}]^{1/2}/2.
\end{align*}
\]

The log-transformed plasma drug concentration given the three consecutive doses is

\[
f_i(V_{il}, k_{a,il}, k_{e,il}, k_{32,il}, k_{23,il}, t) = \log\left\{A_{2,il}(t - t_1) + A_{2,il}(t - t_2) + A_{2,il}(t - t_3)\right\}/V_{il} \}
\]

where \( V_{il} \) is the volume of the distribution for subject \( i \) from dose \( l \). Let \( Y_{ij} \) be the plasma drug concentration for subject \( i \) at the \( j \)th time point (\( j = 1, \cdots, 25 \)). We assume \( Y_{ij} \) follows the nonlinear mixed model

\[
Y_{ij} = f_i(V_{il}, k_{a,il}, k_{e,il}, k_{32,il}, k_{23,il}, t_{ij}) + \epsilon_{ij},
\]

where \( \epsilon_{ij} \sim N(0, \sigma^2) \), and

\[
\begin{align*}
\log(k_{a,il}) &= \log(k_{a,i}) + b_{2i}, & \log(k_{e,il}) &= \log(k_{e,i}) + b_{3i}, \\
\log(k_{23,il}) &= \log(k_{23,i}) + b_{4i}, & \log(k_{32,il}) &= \log(k_{32,i}), \\
\log(V_{il}) &= \log(V_i) + b_{1i},
\end{align*}
\]

and \( b_i = (b_{1i}, b_{2i}, b_{3i}, b_{4i})^T \sim N(0, \mathbf{D}) \), \( \mathbf{D} = \text{diag}(\sigma^2_{11}, \cdots, \sigma^2_{44}) \), \( b_i \) and \( \epsilon_{ij} \) are independent.

We fit the two compartment nonlinear mixed model (2)-(5) using the conditional first order method (Lindstrom and Bates, 1990). Table 1 gives the parameter estimates and their standard errors, and the last column gives the p-values testing for the equivalence of the PK parameters among the three doses. The results show that the absorption rate \( k_{a,i} \) and the distribution rate \( k_{23,i} \) are statistically significantly different among the three doses. The absorption rate decreases with the dose level. This might be explained by the saturability of transporting passages across gastrointestinal membranes. The distribution rate appears to have a bell-shape, with the lowest rate given by the second dose. It is of scientific interest to investigate the factors that drive the bell-shaped relationship between the distribution rate and the dose.
The distribution rate is usually either blood flow sensitive or binding sensitive. It is known in vitro that oxybutynin demonstrates high binding specificity for M3/m3 receptors. If oxybutynin were binding sensitive due to saturation with repeated doses, one would expect the distribution rate decreases monotonically with the doses instead of a bell-shape. In view of our results in Table 1, we have no evidence that oxybutynin is binding sensitive. We hence focus on investigating the factors that might drive the distribution rate to be blood flow sensitive. It is known that blood pressure is a driving force for blood to flow through the arterial system (Eugene, 2001). If the distribution rate is flow sensitive, it could be correlated with blood pressure. In addition, since blood pressure usually has a circadian pattern, it might explain the observed bell-shaped relationship between the distribution rate and the dose. Therefore, we are interested in extending model (2)-(5) to allow for the distribution rate to be a function of blood pressure and heart rate. In the Ditropan study, this investigation is further challenged by the fact that blood pressure is measured with considerably error and its measurement times are different from those of plasma concentration measures. We hence to need take these features into account in our statistical model.

1.3 The Statistical Background

Nonlinear mixed models have been widely used in modeling pharmacokinetic data (Davidian and Giltinan, 1995; Lindstrom and Bates, 1990), where the PK parameters are modeled as parametric functions of time-independent covariates. Li et al. (2002) extended such models to allow for some time-dependent PK parameters to be nonparametric functions of time using splines. Although there is an extensive literature on measurement error in nonlinear models (Carroll et al., 1995), the literature on measurement error in nonlinear mixed effects PK models is rather limited. Higgins, et al. (1997) proposed a simple two-step regression calibration method. They assumed the PK parameters to be time-dependent. Under this assumption, the solution of the differential equation does not have an analytic form. They...
assumed in their estimation procedure an approximate analytic solution. We will show in our simulation study that such a closed-form approximation can be subject to considerable bias. Their two-step method also does not consider the contribution of the variation of the estimator obtained from the first step to the estimation at the second step. Their simulation results show that the model-based standard errors seriously underestimate the true SEs.

We consider in this paper nonlinear mixed effects PK models where the PK parameters are functions of time-dependent covariates which are measured with errors. We describe the model in Section 2, and propose in Section 3 two two-step estimation methods: the second order two-step method with numerical solutions of differential equations (2orderND), and the second order two-step method with closed form approximate solutions of differential equations (2orderAD). Both methods are approximate MLEs where the likelihood function is approximated using the Solomon-Cox-Laplace approximation. They are both computationally easy and stable, and require fitting a linear mixed model at the first step and a nonlinear mixed model at the second step. The proposed methods are applied to the Ditropan study in Section 4, and their performance is evaluated using a simulation study in Section 5. Conclusions are given in Section 6.

2 The Population PK Model with Time-Dependent Covariates Measured with Errors

The preliminary results of the Ditropan data in Table 1 provide no evidence that $V_i$, $k_{e,1}$ and $k_{32,1}$ are dose-dependent, while there is a strong statistical evidence that $k_{a,1}$ and $k_{23,1}$ are dose-dependent. Little between-subject variation of $k_e$ and $k_{32}$ was found. We hence assume $V_i$, $k_{e,1}$ and $k_{32,1}$ are dose-independent and $(k_e, k_{32})$ are fixed effects in our subsequent measurement error PK model. Denote these PK parameters by $\beta_i(t) = \{V_i, k_{a,1}(t), k_e, k_{23,1}(t), k_{32}\}^T$. We consider a two-compartment model similar to (1) except that the PK parameters ($V_{i,l}$, $k_{e,il}$,
\(k_{32,ii}, k_{a,ii}, k_{23,ii}\) are replaced by \(\{V_i, k_e, k_{32}, k_{a,i}(t), k_{23,i}(t)\}\), where \(k_{a,i}(t)\) and \(k_{23,i}(t)\) are time-dependent,

\[
\begin{align*}
\frac{dA_{1,i}(t - t_i)_+}{dt} &= -k_{a,i}(t)A_{1,ii}(t - t_i)_+ \\
\frac{dA_{2,i}(t - t_i)_+}{dt} &= k_{a,i}(t)A_{1,ii}(t - t_i)_+ - \{k_e + k_{23,i}(t)\}A_{2,ii}(t - t_i)_+ \\
&+ k_{32}A_{3,ii}(t - t_i)_+ \\
\frac{dA_{3,i}(t - t_i)_+}{dt} &= k_{23,i}(t)A_{2,ii}(t - t_i)_+ - k_{32}A_{3,ii}(t - t_i)_+,
\end{align*}
\]

(6)

It follows that the differential equations (6) do not have an analytic closed-form solution. We solve it numerically using an IMSL Fortran77 function (DIVPK), which uses the Runge-Kutta-Verner fifth-order method (IMSL USER’s MANUAL STAT/LIBRARY Version 1.0, 1987).

We describe below a three-stage nonlinear mixed model to account for measurement errors in covariates for the two-compartment model. Suppose the data are composed of \(m\) subjects. The \(j\)th observation \((j = 1, \cdots, J_i)\) of the \(i\)th subject \((i = 1, \cdots, m)\) is measured at time \(t_{ij}\) and consists of an outcome variable \(Y_{ij}\), accurately measured time-varying covariates \(X_{ij} = X(t_{ij})\) and an unobserved time-varying covariate \(U_{ij} = U_i(t_{ij})\), e.g., \(U_{ij}\) is the true diastolic blood pressure measured at time \(t_{ij}\) for subject \(i\). Suppose that \(W\) is an observed error-prone measure of \(U\) and is measured at time points \(t_{ik}^* (k = 1, \cdots, K_i)\), where \(t_{ik}^*\) might differ from \(t_{ij}\) and \(J_i\) might differ from \(K_i\). For example, \(W_{ik} = W_i(t_{ik}^*)\) is the observed diastolic blood pressure measured at time \(t_{ik}^*\). In the Ditropan data, the times diastolic blood pressure was taken \((t_{ik}^*)\) differed from the the times plasma concentration was measured \((t_{ij})\), and \(J_i = 25\) and \(K_i = 9\).

The first-stage model describes the observational level pharmacokinetics. Specifically, we rewrite the nonlinear mixed model (5) for the plasma concentration \(Y_{ij}\) \((i = 1, \cdots, m, j = 1, \cdots, J_i)\) as

\[
Y_{ij} = f_i(\beta_{ij}, t_{ij}) + \epsilon_{ij},
\]

(7)
where \( f_i(\beta_{ij}, t_{ij}) = \log \{ A_{2,11}(\beta_{ij}, t_{ij} - t_1) + A_{2,12}(\beta_{ij}, t_{ij} - t_2) + A_{2,13}(\beta_{ij}, t_{ij} - t_3) \}/V_i \), \( A_{2,ii}(\cdot) \) solves (6), \( \beta_{ij} = \beta_i(t_{ij}) \), the \( \epsilon_{ij} \) are independent and follow \( N(0, \sigma^2_\epsilon) \), and are independent of \( \beta_{ij} \).

The second stage model describes the association between time-dependent PK parameter vector \( \beta_{ij} \) and the observed accurately measured covariates \( X_{ij} \) and the unobserved covariate \( U_{ij} \), e.g., \( U_{ij} \) is the true DBP at \( t_{ij} \), as follows

\[
\beta_{ij} = d(X_{ij}, U_{ij}, \alpha, b_i), \quad (8)
\]

where \( \beta_{ij} = \{ V_i, k_{a,i}(t_{ij}), k_e, k_{23,i}(t_{ij}), k_{32} \}^T \), \( V_i = \exp(\alpha_1 + b_{i1}) \), \( k_{a,i}(t_{ij}) = \exp(I(t_{ij} < 8)\alpha_2 + I(8 < t_{ij} < 16)\alpha_3 + I(16 < t_{ij})\alpha_4 + b_{i2}) \), \( I(\cdot) \) is an indicator function, \( k_e = \exp(\alpha_5) \), \( k_{23,i}(t_{ij}) = \{ \alpha_6 + \alpha_7 U_{ij} \} \exp(b_{3i}) \), \( k_{32} = \exp(\alpha_8) \), \( d \) is a 5-dimensional function, \( b_i = (b_{i1}, b_{2i}, b_{3i})^T \sim N(0, D) \), and \( D = \text{diag}(\sigma^2_{11}, \sigma^2_{22}, \sigma^2_{33}) \).

The unobserved covariate \( U_{ij} \) can be regarded as a realization of the underlying true continuous process \( U_i(t) \) at time \( t_{ij} \), i.e., \( U_{ij} = U_i(t_{ij}) \). Suppose \( U_i(t) \) is measured with error by the error-prone process \( W_i(t) \), and \( W_{ik} \) is the value of \( W_i(t) \) at time \( t_{ik}^* \) \( (k = 1, \cdots, K_i) \).

The third stage relates the error-prone covariate \( W_{ik} = W_i(t_{ik}^*) \) to the true unobserved values \( U_i(t_{ik}^*) \) using an additive measurement error model (Carroll et al., 1995)

\[
W_{ik} = U_i(t_{ik}^*) + e_{ik}, \quad (9)
\]

where \( e_{ik} \sim N(0, \sigma^2_e) \) is an independent measurement error and is independent of \( \epsilon_{ij} \) and \( b_i \).

Note that we allow \( t_{ij} \) and \( J_i \) to differ from \( t_{ik}^* \) and \( K_i \). Following Higgins et al. (1997), we assume a polynomial model for the underlying process \( U_i(t_{ik}^*) \)

\[
U(t_{ik}^*) = T_{ik}^T \gamma + B_{ik}^T a_i, \quad (10)
\]

where \( T_{ik} = (1, \cdots, \{ t_{ik}^* \})^T \), \( \gamma \) is a \( (r + 1) \times 1 \) vector, \( B_{ik} = (1, \cdots, \{ t_{ik}^* \})^T \) and \( a_i \) is an \( (h + 1) \times 1 \) vector of random effects following \( N(0, \Omega) \). It follows from (9) and (10) that

\[
W_{ik} = T_{ik}^T \gamma + B_{ik}^T a_i + e_{ik}, \quad (11)
\]
Denote by $Y_i = (Y_{i1}, \cdots, Y_{id})^T$, $Y = (Y_i^T, \cdots, Y_m^T)^T$, and $f$, $t$, $\epsilon$, $\beta$, $X$, $t$, $U$, $t^*$, and $e$ similarly. Let $B = \text{diag}(B_i)$. Equations (7)-(11) can be rewritten in a matrix form

$$
Y = f(\beta, t) + \epsilon, \quad (12)
$$

$$
\beta = d(X, U(t), \alpha, b) = d(X, \gamma, \alpha, b), \quad (13)
$$

$$
W = U(t^*) + e, \quad (14)
$$

$$
U(t^*) = T\gamma + Ba, \quad (15)
$$

where $b \sim N\{0, \text{diag}(D)\}$, $a \sim N\{0, \text{diag}(\Omega)\}$, $\epsilon \sim N(0, \sigma^2_{\epsilon}I)$, and $e \sim N(0, \sigma^2_{\epsilon}I)$, and $b$, $a$, $\epsilon$ and $e$ are independent.

Denote by $\zeta = (\alpha, D, \sigma^2_{\epsilon})$. The integrated likelihood function of the observed data $Y|W, X)$ is

$$
L(Y|W, X; \zeta, \gamma, \Omega, \sigma^2_{\epsilon}) = \int L_1(Y|b, a; \gamma, \zeta)L_2(b; D)L_3(a|W; \gamma, \Omega, \sigma^2_{\epsilon})dbda, \quad (16)
$$

where $L_1(Y|b, a; \gamma, \zeta)$ is the conditional likelihood of $Y$ under the first stage model (12), $L_2(b; D)$ is the likelihood of $b$ under the second stage model (13), and $L_3(a|W; \gamma, \Omega, \sigma^2_{\epsilon})$ is the likelihood of $a$ under the third stage models (14)-(15). Since the function $f(\cdot)$ is nonlinear, the likelihood (16) does not have a closed form and its evaluation involves a high dimensional integral.

## 3 The Two Estimation Procedures

### 3.1 The Solomon-Cox-Laplace Approximation of the Likelihood

In view of the high dimensional integral involved in the likelihood (16), we consider a two-step estimation procedure by approximating the likelihood (16) using the Solomon-Cox-Laplace approximation. Since the $W$ data contain majority of the information about $(\gamma, \Omega, \sigma^2_{\epsilon})$, at the first step, we use the $W$ data to estimate $(\gamma, \Omega, \sigma^2_{\epsilon})$ by fitting the linear mixed model (11)
and obtain the BLUP estimator of $\xi = (\gamma^T, a^T)$ denoted by $\tilde{\xi}$ and its covariance $\tilde{V}_\xi$. It can be easily shown that the resulting estimators $(\tilde{\gamma}, \tilde{\Omega}, \tilde{\sigma}_e^2)$ are consistent. At the second step, we treat $\tilde{\xi}$ and $\tilde{V}_\xi$ to be known and maximize the Solomon-Cox-Laplace approximation of the integrated likelihood (16) as a function of $\zeta$ to obtain an approximate MLE of $\zeta$.

Specifically, denote by $U = U(t) = T\gamma + Ba = S\xi$, where $S = (T, B)$ and $\xi = (\gamma^T, a^T)^T$. Given $(\tilde{\gamma}, \tilde{\Omega}, \tilde{\sigma}_e^2)$, the distribution of $(U|W)$ can be estimated using the distribution of $(\xi|W)$, i.e., the distribution of the BLUP estimator of $\xi$. Denote by $H = SS^T + \text{diag}(0, \tilde{\sigma}_e^2\tilde{\Omega}^{-1})$. Then $\xi|W$ follows $N(\tilde{\xi}, \tilde{V}_\xi)$, where

$$\tilde{\xi} = H^{-1}SW, \quad \tilde{V}_\xi = \tilde{\sigma}_e^2H^{-1}. \tag{17}$$

It follows that the distribution of $U|W$ can estimated as $N(\overline{U}, \overline{V}_u)$, where $\overline{U} = S\tilde{\xi}$ and $\overline{V}_u = SS\tilde{\xi}S^T$. Note that in fact $\overline{U} = E(U|W)$ evaluated at $(\tilde{\gamma}, \tilde{\Omega}, \tilde{\sigma}_e^2)$, and $\overline{U}$ corresponds to the regression calibration estimator of $U$.

To estimate $\zeta$, we approximate the doubly integrated likelihood (16) using the Solomon-Cox and Laplace approximations at the second stage. Specifically, some calculations show that the likelihood (16) can be rewritten by integrating out $U$ and $b$ as

$$L(Y|W, X; \zeta, \gamma, \Omega, \sigma_e^2) = \int L_1(Y|b, U; \gamma) L_2(b; D) L_3(U|W; \gamma, \Omega, \sigma_e^2) dU db, \tag{18}$$

where $L_3(U|W)$ is the likelihood of $U|W$ and can be estimated as $U|W \sim N(\overline{U}, \overline{V}_u)$. To approximate the double integrals in (18), we first approximate the integral that involves $U$ using the Solomon-Cox approximation (Solomon and Cox, 1992) by treating the integrand as a function of $U$ and taking a quadratic expansion of the integrand about $\overline{U}$, the mean of $U$. We next approximate the integral that involves $b$ using the Laplace approximation by treating the integrand as a function of $b$ and taking a quadratic expansion of the integrand about its mode $\tilde{b}$. The resulting Solomon-Cox-Laplace approximation can be regarded as the second order approximation of the integrated likelihood (16).

Specifically, following Wolfinger and Lin (1997, Section 3), one can show that the Solomon-Cox approximation for the integral involving $U$ corresponds to the loglikelihood of $Y$ as a
function of $\zeta$ under the nonlinear mixed model

$$Y = f(\beta, t) + e^*, \quad (19)$$

where $\beta = d(X, \bar{U}, \alpha, b) = d(X, \tilde{\zeta}, \alpha, b), \ b \sim N(0, D), \ e^* \sim N(0, R^*), \ R^* = \sigma^2 \mathbf{I} + \mathbf{F}_u \bar{V}_u \mathbf{F}_u^T = \sigma^2 \mathbf{I} + \mathbf{F}_\zeta \mathbf{V}_\zeta^{-1} \mathbf{F}_\zeta^T$, and $\mathbf{F}_u = \partial f/\partial \bar{U}^T$ and $\mathbf{F}_\zeta = \partial f/\partial \zeta^T$. Note that here we assume $(\bar{U}, \bar{V}_u)$, equivalently $(\tilde{\zeta}, \bar{V}_\zeta)$, which are obtained from the first step, are fixed and known parameters.

Following Lindstrom and Bates (1990) and Wolfinger and Lin (1997, Section 4), we can show that a further application of the Laplace approximation for the integral involving $b$ in (18) gives, apart from a constant,

$$\ell_{SCL}(Y|W; \zeta) = -\frac{1}{2} \ln |\bar{V}| - \frac{1}{2} \left\{ Y - f(\tilde{\beta}, t) - \bar{Z}\tilde{b} \right\} \bar{V}^{-1} \left\{ Y - f(\tilde{\beta}, t) - \bar{Z}\tilde{b} \right\}, \quad (20)$$

where $\tilde{\beta} = d(X, \bar{U}, \alpha, \tilde{b}), \ \bar{V} = \bar{Z}D\bar{Z}^T + R^*, \ \bar{Z} = \partial f/\partial b^T|_{b=\tilde{b}}$ and $\tilde{b}$ solves

$$\frac{1}{\sigma^2_\zeta} \bar{Z}^T \left\{ Y - f(\tilde{\beta}, t) \right\} - D^{-1}\tilde{b} = 0.$$

We term (20) the Solomon-Cox-Laplace approximation of the likelihood (16). The resulting approximatie maximum likelihood estimator of $\zeta$ that maximizes (20) corresponds to fitting the nonlinear mixed model (19) using the Lindstrom and Bates’ (1990) algorithm.

3.2 The Two-Stage Estimation Methods

The above discussions suggest that our two-step estimation procedure is hence simple. At the first step, one fits the linear mixed model (11) using the $W$ data to obtain the MLE ($\tilde{\gamma}, \tilde{\Omega}, \tilde{\sigma}^2$) and the BLUP estimator $\tilde{\zeta}$ and its covariance $\bar{V}_\zeta$. At the second step, one treats $\tilde{\zeta}$ and $\bar{V}_\zeta$ as known, and fits the nonlinear mixed model (19) with $b$ as the random effects using the Lindstrom and Bates algorithm.

Following Lindstrom and Bates (1990) and Wolfinger and Lin (1997), one can show that fitting the nonlinear mixed model at the second stage can proceed by iteratively fitting the
linear mixed model

\[ Y^* = X^* \alpha + Z^* b + e^*, \]  

(21)

where denoting by \( \alpha^* \) and \( b^* \) the estimates at the previous iteration, \( Y^* \) is a working vector,

\[ Y^* = Y - f^* + X^* \alpha + Z^* b^*, \quad e^* \sim N(0, R^* = F_\xi \bar{V}_\xi F_\xi^T) \]

\[ f^* = \frac{\partial f}{\partial \alpha^T} |_{\alpha^*, b^*}, \quad X^* = \frac{\partial f}{\partial \alpha^T} |_{\alpha^*, b^*}, \xi \]

\[ Z^* = \frac{\partial f}{\partial b^T} |_{\alpha^*, b^*}, \xi , \quad F_\xi = \frac{\partial f}{\partial \xi^T} |_{\alpha^*, b^*}, \xi . \]

The variance components \( D \) and \( \sigma^2_\epsilon \) are estimated iteratively using REML under (21).

Note that the regression calibration method (Higgins et al., 1997) ignores the variability of \( U|W \) by simply plugging \( \bar{U} \) into \( \beta(\cdot) \) and setting \( U = \bar{U} \). Our procedure accounts for the variability in the regression calibration estimator \( \bar{U} \).

The nonlinear mixed model (19) and the linear mixed model (21) involve evaluating the nonlinear function \( f(\cdot) \). We propose two two-step estimation procedures in the next two sections. Both procedures are the same at the first step. They differ by the second step. The first procedure evaluates \( f(\cdot) \) by numerically solving the differential equation (6) at the second step, while the second one approximates the solution of (6) analytically.

3.3 The Second-Order Two-Step Method with Numerical Solution of Differential Equations

Our first two-step estimation procedure fits the linear mixed model (11) using the \( W \) data at the first step and fits the nonlinear mixed model (19) or iteratively fits the linear mixed model (21), where \( f(\cdot) \) is evaluated by numerically solving the differential equation (6). We call this method the second-order two-step method with numerical solution of differential equations (2orderND).

Examination of equation (21) suggests that we need to calculate both \( f_\{\beta(t), t\} \) and its first derivative \( \partial f_\{\beta(t), t\}/\partial \beta \). For the two compartment model, equation (6) indicates that
we need to calculate $A_{ii}(t-t_i)_+$ and $A_{i,i}(t-t_i)_+ = \partial A_{ii}(t-t_i)_+/\partial \beta$, where $A_{ii}(t-t_i)_+ = \{A_{1,ii}(t-t_i)_+, A_{2,ii}(t-t_i)_+, A_{3,ii}(t-t_i)_+ \}^T$. Write (6) as $A_{ii}(t-t_i)_+/dt = G_i\{\beta(t)\} A_{ii}(t-t_i)_+$ and take a derivative with respect to $\beta$ on both sides, we have

$$\frac{dA_{i,i}(t-t_i)_+}{dt} = G_{i,i}\{\beta(t)\} A_{ii}(t-t_i)_+ + G_i\{\beta(t)\} A_{i,i}(t-t_i)_+,$$

(22)

where $G_i(:, \beta) = \partial G_i(\cdot)/\partial \beta$. Hence both $A_{ii}(\cdot)$ and $A_{i,i}(\cdot)$ can be obtained by solving the differential equations (1) and (22). Note that the initial value $A_{i,i}(0)$ is always 0, because changing the PK parameters $\beta$ will not affect the initial value $A_{ii}(0)$. More details of the numerical solution of the differential equations can be found in Li et al. (2002).

Wu (2002) applied an EM algorithm to maximize a marginal likelihood function similar to (18). He treated both the random effects $b$ and the unobserved variable $U$ as missing data. At the E-step, $b$ and $U$ were estimated using the observed data $(Y, W)$ by assuming the conditional distribution of $(b, U|Y, W)$ to be normal. At the M-step, given the estimated $b$ and $U$, he maximized the joint likelihood function of $(Y, W, b, U)$ over all the other parameters.

Our two-step method assumes that almost all of the information about $U$ is contained in $W$. Hence, $U$ is estimated from $W$ exclusively at step one. At step two, the parameters of the PK model and the random effects $b$ are estimated jointly from the Solomon-Cox-Laplace approximation (20) of the likelihood function of $Y$. Since he assumed normality for the conditional distribution of $(b, U|Y, W)$, his EM algorithm can be viewed as a second order approximation of the marginal likelihood function (16), similar to ours. However, unlike his method, we estimate $U$ only once, and therefore the computational burden required by numerically solving the differential equations is considerably reduced.
3.4 Second Order Two-Step Method with Approximately Closed Form Solutions of Differential Equations

Unless the PK parameters are time-independent, there is no closed form solution for the differential equations (6) in general. However, if the PK parameters change only slightly over time, the closed form solution based on the constant PK parameter assumption would provide a good approximation to the exact solution. In this section, We make such an assumption and solve the differential equations analytically in a closed form. The resulting estimation procedure hence is computationally much faster. Specifically, under this assumption, the solution of (6) is

\[
A_{2,ii}(t - t_i) = \frac{k_{a,i}(t)\{k_{32} - k_{a,i}(t)\}e^{-k_{a,i}(t)(t-t_i)_+}}{\lambda_{1,i}(t) - k_{a,i}(t)\{\lambda_{2,i}(t) - k_{a,i}(t)\}} + \frac{k_{a,i}(t)\{k_{32} - \lambda_{1,i}(t)\}e^{-\lambda_{1,i}(t)(t-t_i)_+}}{\lambda_{1,i}(t) - \lambda_{2,i}(t)\{k_{a,i}(t) - \lambda_{2,i}(t)\}}
\]

(23)

where

\[
\lambda_{1,i}(t) = \left[ k_c + k_{23,i}(t) + k_{32} + \{(k_c + k_{23,i}(t) + k_{32})^2 - 4k_c k_{32}\}^{1/2} \right] / 2,
\]

\[
\lambda_{2,i}(t) = \left[ k_c + k_{23,i}(t) + k_{32} - \{(k_c + k_{23,i}(t) + k_{32})^2 - 4k_c k_{32}\}^{1/2} \right].
\]

We call this method as the second order two-step method with approximately closed form solutions of differential equations (2orderAD). It is the same as the 2orderND method described in Section 3 except that a closed form expression \( f(\cdot) \) is used in the estimation at the second step when fitting (19).

4 Data Analysis

We applied the proposed measurement error PK model (7)-(10) to the analysis of the Ditropan data introduced in Section 1 to study the effect of DBP on the distribution rate
\( k_{23}(t) \). We also considered the effects of SBP and heart rate (HR) on \( k_{23}(t) \). Since DBP, SBP and HR are highly correlated, we analyzed each of them separately by assuming \( W \) to be either of these three variables.

At step one, the linear mixed model (10) with a random intercept was fitted to DBP, SBP and heart rate separately using the Splus function LME. We modeled the time effects using quadratic, cubic and quartic regression, and compared their fits using the AICs. The quartic model for DBP, the cubic models for SBP and heart rate gave the lowest AIC. The AIC values of all models and the parameter estimates associated with the model with the smallest AICs for each variable are given in Table 2. Higher order random effects models, e.g., random slope models, were also tried. They all had higher AIC values. The solid lines in Figure 1 (a), (b) and (c) give the polynomial fits with the lowest AICs for these three time-dependent covariates. The curves all first decreased, then increased, and then decreased again. This indicates that blood pressure and heart rate react to the three doses.

At step two, the plasma concentration was modeled using the nonlinear mixed model (19), where \( f_i(\cdot) = f[A_2,i_1(t - t_1) + A_2,i_2(t - t_2) + A_2,i_3(t - t_3)] \), \( A_2,i(t) \) is given (6), and

\[
k_{23,i}(t_{ij}) = \{\alpha_6 + \alpha_7 U(t_{ij})\} \exp(b_k),
\]

and \( U(t) \) is set to be either DBP, SBP or HR. Parameter estimators were obtained using the 2orderND method and the 2orderAD method described in Section 3 to account for measurement error in DBP, SBP and HR. The results are given in Table 3. Using the 2orderND method, we found that \( k_{23,i}(t) \) was significantly positively associated with true \( DBP(t) \) (\( \alpha_7 = 0.0042, p\text{-value}=0.022 \)), and there was some evidence of a positive association between \( k_{23,i}(t) \) and true \( SBP(t) \) (\( \alpha_7 = 0.0023, p\text{-value} = 0.125 \)) and true \( HR(t) \) (\( \alpha_7 = 0.0032, p\text{-value} = 0.075 \)). These results partially support our hypothesis that the distribution process could be blood flow sensitive.

To investigate the effect of not incorporating the time-varying covariate, e.g., DBP, in the \( k_{23,i}(t) \) model, we compared the residuals of the model assuming \( k_{23,i}(t) \) was a function
of DBP given in equation (24) with \( U(t) = \text{true DBP}(t) \), with the model assuming \( k_{23,i}(t) \) was free of DBP as \( k_{23,i}(t) = \alpha_0\exp(b_{3i}) \). These two residual plots are given in Figure 2(a) and (b). Figure 2(c) shows the average differences between these two sets of residuals, and Figure 2 (d) shows the estimate curve of \( k_{23}(t) \) as a function of \( t \) and its 95% confidence interval.

Figure 2(c) suggests that if the effect of DBP is not modeled, the residuals are too high during the second dosing period, and too low during the first and the third dosing periods. In other words, there was a clear lack of fit if the mean of \( k_{23,i}(t) \) was assumed to be a constant and free of DBP. This lack of fit was caused by the fact that the second dose had a lower \( k_{23} \) than the other two doses, and the lower \( k_{23} \) in the second dosing period might be due to the fact that a higher DBP was associated with the second dose.

We also analyzed the data using the 2orderAD method, where model (24) was used in calculating the mean function \( f(\cdot) \) in (4). The results in Table 3 show that the PK parameters were overestimated using the 2orderAD method. Our simulation study confirms this finding. We present 2orderAD analysis with only DBP in Table 3, the other two time-dependent covariates have the similar results. The biases of the 2orderAD estimates were likely to result from the poor approximation of the closed form solution of the differential equations. Both methods however provided comparable variance component estimates.

5 Simulation

We performed a simulation study to evaluate the performance of the two-step 2orderND and 2orderAD procedures. The design of the simulation study was identical to that of the original Ditropan data. Specifically, we generated the data from model (7)-(10) with \( f(\cdot) \) given in (4), and set the true parameter values equal to the estimates from the analysis of the real DBP data given in Tables 2 and 3. In each simulated data set, there were 40 subjects and each subject had 25 observations over time. In addition, every subject’s DBP
was simulated at 9 different fixed sampling time points during 48 hours. We used 500 Monte Carlo data sets.

We analyzed each simulated data set using the 2orderND and 2orderAD methods. At step one, the AIC was used to estimate the degree of the polynomial of $DBP(t)$. Hence both methods had the same estimate of true $DBP(t)$. At step two, PK parameters were estimated from the mean function which was calculated using the differential equations (2orderND) or the approximate analytic solution of the differential equations (2orderAD). Table 4 summarizes the average point estimates, model-based SEs (MOD-SE), empirical SEs (EMP-SE), relative biases of the parameter estimates, and the ratios of the model-based SEs and the empirical SEs.

When the 2orderND method was used, 6 out of 500 data sets did not converge. The results in Table 4 show that the 2orderND method performs well. The PK parameter estimates had small biases, with the largest 12% bias observed in the estimate of $\alpha_7$, which describes the association between true $DBP(t)$ and $k_{23}(t)$. The variance component estimates also had small biases. The biases of the estimates of $\sigma_\epsilon^2$ and $\sigma_{11}^2$ were smaller than those of $\sigma_{22}^2$ and $\sigma_{33}^2$. These results are expected, as $\epsilon_{ij}$ and $b_{1i}$ are linearly related to $y_{ij}$, while $b_{2i}$ and $b_{3i}$ are nonlinearly related to $y_{ij}$. The model based SEs slightly underestimated the true SEs.

The 2orderAD method was slightly less stable than the 2orderND method, and 9/500 of the simulated data sets did not converge. The results in Table 4(b) show that the estimates are considerably biased especially the estimates of $\alpha_6$ and $\alpha_7$, whose biases could be as high as 80%. This is expected, since the 2orderAD method uses an approximate analytic solution of the differential equations by assuming the PK parameters change slowly with $t$. If the PK parameters vary considerably with the time varying covariate, the approximation is likely to be poor. Both methods however provided comparable variance component estimates.
6 Conclusion

We considered in this paper a population PK model with the PK parameters modeled as functions of time dependent covariates measured with errors. We proposed two two-step estimation methods. Both methods are computationally easy and stable, and only require fitting a linear mixed model at the first stage followed by fitting a nonlinear mixed model at the second stage. The 2orderND method accounts for the fact that the PK parameters depend on a time varying covariate and change over time, and solves the differential equations numerically; while the 2orderAD method assumes the PK parameters vary slowly with time and approximates the solution of the differential equations in a closed form. Our simulation results show that the 2orderND method performs well, and has small biases in parameter estimators and the model based standards errors are close to the empirical standard errors. The 2orderAD method is subject to considerable bias. For simplicity, our presentation focused on the two-compartment model in this paper. The proposed method is applicable to general nonlinear mixed models commonly used in pharmacokinetics.

We showed that the 2orderND method could be derived as the Solomon-Cox-Laplace approximation of the integrated likelihood. Our simulation study suggests that such a likelihood approximation works well in practice. In view of the computational advantage of the proposed method and the considerable additional computational burden of the maximum likelihood method, it would be of future research interest to investigate whether the magnitude of improvement of the maximum likelihood method over the 2orderND method is practically appreciable and is worth additional computational burden.

Based on our proposed 2orderND method, we found using the Ditropan data that the true underlying processes of diastolic blood pressure, systolic blood pressure and heart rate were all positively correlated with $k_{23}(t)$, although only diastolic blood pressure showed a statistically significant association. This finding suggests that drug distribution rate might be flow sensitive as discussed at the end of Section 1.2. To have a better power to verify the
association between $k_{23}(t)$ and blood pressure and heart rate, a larger range of systematically changed blood pressure and heart rate is needed. For example, it would be desirable to let subjects exercise during one dosing period, and rest in another period in a new study. On the other hand, in order to understand the association between blood pressure and S-oxynbutynin’s pharmacokinetics among hypertension and hypotension patients, more studies are necessary. As this study was performed on healthy subjects, it remains for further studies to verify our findings and determine its clinical significance in the patient population. If the decreasing $k_a$ pattern is verified, the controlled release formulation may need to be modified to keep the drug concentration as stable as possible. If the flow sensitive hypothesis of $k_{23}$ is verified, more studies may need to be done carefully among hypertension and hypotension patients.

**Acknowledgment**

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**References**

Bethesda, MD : American Society of Hospital Pharmacists.


Table 1  Parameter Estimates Under the Dose-specific Two-Compartment Model (2)-(5) for the Ditropan Study

<table>
<thead>
<tr>
<th>Fixed Effects</th>
<th>dose 1</th>
<th>dose 2</th>
<th>dose 3</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>$V$</td>
<td>0.163 (0.021)</td>
<td>0.175 (0.020)</td>
<td>0.195 (0.013)</td>
<td>0.687</td>
</tr>
<tr>
<td>$k_a$</td>
<td>6.394 (0.882)</td>
<td>3.553 (0.391)</td>
<td>2.574 (0.207)</td>
<td>0.001</td>
</tr>
<tr>
<td>$k_c$</td>
<td>0.652 (0.027)</td>
<td>0.605 (0.026)</td>
<td>0.632 (0.022)</td>
<td>0.330</td>
</tr>
<tr>
<td>$k_{23}$</td>
<td>0.312 (0.060)</td>
<td>0.209 (0.051)</td>
<td>0.324 (0.015)</td>
<td>0.041</td>
</tr>
<tr>
<td>$k_{32}$</td>
<td>0.122 (0.104)</td>
<td>0.078 (0.082)</td>
<td>0.091 (0.025)</td>
<td>0.914</td>
</tr>
</tbody>
</table>

Variance Components

<p>| | |</p>
<table>
<thead>
<tr>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>$\sigma^2_e$</td>
<td>0.135 (0.011)</td>
</tr>
<tr>
<td>$\sigma_{11}$</td>
<td>0.247 (0.039)</td>
</tr>
<tr>
<td>$\sigma_{22}$</td>
<td>0.162 (0.046)</td>
</tr>
<tr>
<td>$\sigma_{33}$</td>
<td>0.121 (0.094)</td>
</tr>
<tr>
<td>$\sigma_{44}$</td>
<td>0.204 (0.095)</td>
</tr>
</tbody>
</table>
Table 2 AICs and Parameter Estimates of the Polynomial Mixed Models (10) for DBP, SBP and HR for the Ditropan Study

<table>
<thead>
<tr>
<th></th>
<th>Diastolic Blood Pressure</th>
<th>Systolic Blood Pressure</th>
<th>Heart Rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>AIC</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Linear</td>
<td>1844.2</td>
<td>1874.0</td>
<td>1857.9</td>
</tr>
<tr>
<td>Quadratic</td>
<td>1839.5</td>
<td>1871.4</td>
<td>1854.2</td>
</tr>
<tr>
<td>Cubic</td>
<td>1827.1</td>
<td>1868.4</td>
<td>1850.4</td>
</tr>
<tr>
<td>Quartic</td>
<td>1826.8</td>
<td>1879.9</td>
<td>1855.5</td>
</tr>
<tr>
<td>Parameter Estimate&lt;sup&gt;a&lt;/sup&gt;</td>
<td>Intercept</td>
<td>66.76</td>
<td>107.97</td>
</tr>
<tr>
<td></td>
<td>(t − 13)</td>
<td>0.33</td>
<td>0.11</td>
</tr>
<tr>
<td></td>
<td>(t − 13)&lt;sup&gt;2&lt;/sup&gt;</td>
<td>0.02</td>
<td>0.02</td>
</tr>
<tr>
<td></td>
<td>(t − 13)&lt;sup&gt;3&lt;/sup&gt;</td>
<td>-0.002</td>
<td>-0.0004</td>
</tr>
<tr>
<td></td>
<td>(t − 13)&lt;sup&gt;4&lt;/sup&gt;</td>
<td>0.00004</td>
<td></td>
</tr>
<tr>
<td></td>
<td>$\sigma_a^2$</td>
<td>6.43</td>
<td>8.08</td>
</tr>
<tr>
<td></td>
<td>$\sigma_c^2$</td>
<td>5.90</td>
<td>7.27</td>
</tr>
</tbody>
</table>

<sup>a</sup> The parameter estimates of the model with the smallest AIC.
Table 3 Parameter Estimates Under the PK Model with the Time-Dependent Covariate Measured with Error (7)-(10) for the Ditropan Study

<table>
<thead>
<tr>
<th>Parameters</th>
<th>2orderND (SE) (DBP)</th>
<th>2orderND (SE) (SBP)</th>
<th>2orderND (SE) (HR)</th>
<th>2orderAD (SE) (DBP)</th>
</tr>
</thead>
<tbody>
<tr>
<td>$V = e^{a_1}$</td>
<td>0.171 (0.010)</td>
<td>0.170 (0.010)</td>
<td>0.171 (0.010)</td>
<td>0.187(0.009)</td>
</tr>
<tr>
<td>$k_{a1} = e^{a_2}$</td>
<td>6.536 (0.839)</td>
<td>6.547 (0.854)</td>
<td>6.541 (0.861)</td>
<td>7.232(1.112)</td>
</tr>
<tr>
<td>$k_{a2} = e^{a_3}$</td>
<td>3.591 (0.327)</td>
<td>3.668 (0.344)</td>
<td>3.625 (0.342)</td>
<td>4.654(0.400)</td>
</tr>
<tr>
<td>$k_{a3} = e^{a_4}$</td>
<td>2.612 (0.169)</td>
<td>2.673 (0.173)</td>
<td>2.650 (0.166)</td>
<td>3.276(0.287)</td>
</tr>
<tr>
<td>$k_{c} = e^{a_5}$</td>
<td>0.603 (0.016)</td>
<td>0.611 (0.017)</td>
<td>0.607 (0.016)</td>
<td>0.455(0.020)</td>
</tr>
<tr>
<td>$\alpha_6$</td>
<td>-0.040 (0.088)</td>
<td>-0.013 (0.072)</td>
<td>-0.025 (0.077)</td>
<td>-0.079(0.111)</td>
</tr>
<tr>
<td>$\alpha_7$</td>
<td>0.0042 (0.002)</td>
<td>0.0023 (0.0015)</td>
<td>0.0032 (0.0018)</td>
<td>0.0059 (0.0017)</td>
</tr>
<tr>
<td>$k_{32} = e^{a_8}$</td>
<td>0.091 (0.008)</td>
<td>0.084 (0.008)</td>
<td>0.085 (0.008)</td>
<td>0.075(0.006)</td>
</tr>
<tr>
<td>$\sigma_c^2$</td>
<td>0.162 (0.012)</td>
<td>0.164 (0.012)</td>
<td>0.164 (0.012)</td>
<td>0.166(0.013)</td>
</tr>
<tr>
<td>$\sigma_{11}$</td>
<td>0.265 (0.037)</td>
<td>0.269 (0.038)</td>
<td>0.266 (0.037)</td>
<td>0.255(0.044)</td>
</tr>
<tr>
<td>$\sigma_{22}$</td>
<td>0.167 (0.042)</td>
<td>0.166 (0.042)</td>
<td>0.167 (0.042)</td>
<td>0.170(0.039)</td>
</tr>
<tr>
<td>$\sigma_{33}$</td>
<td>0.256 (0.111)</td>
<td>0.273 (0.122)</td>
<td>0.265 (0.118)</td>
<td>0.268(0.121)</td>
</tr>
</tbody>
</table>

- 2orderND: second order two-step method with the numerical solutions of the differential equations.
- 2orderAD: second order two-step method with the approximately closed form solution of the differential equations.
**Table 4(a) Simulation Results Using the 2orderND Method**

<table>
<thead>
<tr>
<th>Parameter</th>
<th>True Value</th>
<th>Estimate</th>
<th>MOD-SE</th>
<th>EMP-SE</th>
<th>RB</th>
<th>MOD-SE/EMP-SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>$V = e^{a_1}$</td>
<td>0.17</td>
<td>0.165</td>
<td>0.007</td>
<td>0.008</td>
<td>-0.029</td>
<td>0.875</td>
</tr>
<tr>
<td>$k_{a1} = e^{a_2}$</td>
<td>6.50</td>
<td>5.877</td>
<td>0.649</td>
<td>0.711</td>
<td>-0.096</td>
<td>0.913</td>
</tr>
<tr>
<td>$k_{a2} = e^{a_3}$</td>
<td>3.60</td>
<td>3.356</td>
<td>0.219</td>
<td>0.221</td>
<td>-0.065</td>
<td>0.990</td>
</tr>
<tr>
<td>$k_{a3} = e^{a_4}$</td>
<td>2.50</td>
<td>2.457</td>
<td>0.182</td>
<td>0.212</td>
<td>-0.017</td>
<td>0.858</td>
</tr>
<tr>
<td>$k_c = e^{a_5}$</td>
<td>0.60</td>
<td>0.612</td>
<td>0.021</td>
<td>0.022</td>
<td>0.020</td>
<td>0.954</td>
</tr>
<tr>
<td>$\alpha_6$</td>
<td>-0.04</td>
<td>-0.035</td>
<td>0.081</td>
<td>0.091</td>
<td>-0.125</td>
<td>0.890</td>
</tr>
<tr>
<td>$\alpha_7$</td>
<td>0.0042</td>
<td>0.0038</td>
<td>0.0016</td>
<td>0.0018</td>
<td>-0.095</td>
<td>0.888</td>
</tr>
<tr>
<td>$k_{32} = e^{a_8}$</td>
<td>0.09</td>
<td>0.092</td>
<td>0.008</td>
<td>0.009</td>
<td>0.022</td>
<td>0.888</td>
</tr>
<tr>
<td>$\sigma_{e}^2$</td>
<td>0.162</td>
<td>0.174</td>
<td>0.014</td>
<td>0.016</td>
<td>0.074</td>
<td>0.875</td>
</tr>
<tr>
<td>$\sigma_{11}$</td>
<td>0.265</td>
<td>0.247</td>
<td>0.042</td>
<td>0.046</td>
<td>-0.068</td>
<td>0.913</td>
</tr>
<tr>
<td>$\sigma_{22}$</td>
<td>0.167</td>
<td>0.149</td>
<td>0.049</td>
<td>0.059</td>
<td>-0.108</td>
<td>0.830</td>
</tr>
<tr>
<td>$\sigma_{33}$</td>
<td>0.256</td>
<td>0.207</td>
<td>0.152</td>
<td>0.178</td>
<td>-0.191</td>
<td>0.853</td>
</tr>
</tbody>
</table>
### Table 4 (b) Simulation Results Using the 2orderAD Method

<table>
<thead>
<tr>
<th>Parameter</th>
<th>True Value</th>
<th>Estimate</th>
<th>MOD-SE</th>
<th>EMP-SE</th>
<th>RB</th>
<th>MOD-SE/EMP-SE</th>
</tr>
</thead>
<tbody>
<tr>
<td>$V = e^{a_1}$</td>
<td>0.17</td>
<td>0.192</td>
<td>0.007</td>
<td>0.008</td>
<td>0.129</td>
<td>0.875</td>
</tr>
<tr>
<td>$k_{a1} = e^{a_2}$</td>
<td>6.50</td>
<td>8.353</td>
<td>1.013</td>
<td>1.210</td>
<td>0.285</td>
<td>0.837</td>
</tr>
<tr>
<td>$k_{a2} = e^{a_3}$</td>
<td>3.60</td>
<td>4.872</td>
<td>0.364</td>
<td>0.429</td>
<td>0.353</td>
<td>0.848</td>
</tr>
<tr>
<td>$k_{a3} = e^{a_4}$</td>
<td>2.50</td>
<td>3.030</td>
<td>0.271</td>
<td>0.308</td>
<td>0.212</td>
<td>0.879</td>
</tr>
<tr>
<td>$k_c = e^{a_5}$</td>
<td>0.60</td>
<td>0.490</td>
<td>0.018</td>
<td>0.019</td>
<td>0.183</td>
<td>0.947</td>
</tr>
<tr>
<td>$\alpha_6$</td>
<td>-0.040</td>
<td>-0.0068</td>
<td>0.092</td>
<td>0.102</td>
<td>-0.830</td>
<td>0.902</td>
</tr>
<tr>
<td>$\alpha_7$</td>
<td>0.0042</td>
<td>0.0057</td>
<td>0.0018</td>
<td>0.0020</td>
<td>0.357</td>
<td>0.900</td>
</tr>
<tr>
<td>$k_{32} = e^{a_8}$</td>
<td>0.09</td>
<td>0.076</td>
<td>0.006</td>
<td>0.008</td>
<td>-0.156</td>
<td>0.750</td>
</tr>
<tr>
<td>$\sigma^2$</td>
<td>0.162</td>
<td>0.175</td>
<td>0.015</td>
<td>0.018</td>
<td>0.080</td>
<td>0.833</td>
</tr>
<tr>
<td>$\sigma_{11}$</td>
<td>0.265</td>
<td>0.242</td>
<td>0.045</td>
<td>0.049</td>
<td>-0.087</td>
<td>0.917</td>
</tr>
<tr>
<td>$\sigma_{22}$</td>
<td>0.167</td>
<td>0.143</td>
<td>0.054</td>
<td>0.067</td>
<td>-0.144</td>
<td>0.805</td>
</tr>
<tr>
<td>$\sigma_{33}$</td>
<td>0.256</td>
<td>0.201</td>
<td>0.151</td>
<td>0.181</td>
<td>-0.214</td>
<td>0.834</td>
</tr>
</tbody>
</table>

- **2orderND**: second order two-step method with the numerical solutions of the differential equations.
- **2orderAD**: second order two-step method with the approximately closed form solution of the differential equations.
- **MOD-SE**: Model-based standard error.
- **EMP-SE**: Empirical standard error.
- **RB**: Relative bias.
Figure 1: Raw data: (a) diastolic blood pressure (DBP), (b) systolic blood pressure (SBP), (c) heart rate (HR), and (d) S-oxybutynin plasma concentration. The solid lines in (a), (b) and (c) are the fitted polynomial curves corresponding to the lowest AICs using the linear mixed model (11).

Figure 2: (a) The residual plot when $k_{23,i}(t)$ was modeled as $\{a_6 + a_7 \times \text{true}DBP(t)\} \exp(b_3)$; (b) The residual plot when $k_{23,i}(t)$ was modeled as $\{a_6\} \exp(b_3)$; (c) The average residual differences between (a) and (b); (d) The estimate of $k_{23,i}(t)$ as a function of $t$ under (a): Solid line is the fitted curve of $k_{23,i}(t)$; the dotted lines are its 95% confidence interval.
Figure 1:
Figure 2: